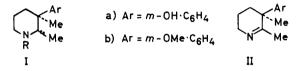
Isomeric 2,3-dimethyl-3-arylpiperidines with morphine-and nalorphine-like properties

Kugita, Oine & others (1965) previously reported that single isomeric forms of the 2,3-dimethyl-3-arylpiperidines Ia ($R = CH_2CH_2Ph$ and CH_2COPh) were about half as active as morphine in mice (hot-plate test) while the corresponding *N*-allyl derivative antagonized the analgesic effects of morphine in the same animal (see also Nurimoto & Hayashi, 1973). We obtained both *RS*-diastereoisomers (termed α - and β -) of the parent secondary amine Ib (R = H) by hydrogenation of the tetrahydropyridine II.



The configurations α : c-2-Me-r-3-Ar and β : t-2-Me-r-3-Ar were established by analysis of differences in the ¹H nmr spectra of isomeric pairs, particularly of N-benzyl and N-acetyl derivatives (Casy & Iorio, 1974). O-Demethylation of the derivatives Ib (R = Me, CH₂CH₂Ph, and CH₂CH:CH₂) was achieved by treating the methoxy compound (1 g) with hydrogen bromide (10 ml, 48% in water) at the reflux temperature for 45 min under nitrogen, and the recovered base converted to a hydrochloride salt. Details of phenolic derivatives so prepared are given in Table 1.

In both hot-plate and Nilsen tests for analgesia in mice (Perrine, Atwell & others, 1972), the *N*-phenethyl pair (Ia, $R = CH_2CH_2Ph$) was significantly active (α : 0.7 ×, β : 0.3 to 0.4 × pethidine) (Table 2), while *N*-methyl and *N*-allyl analogues were very weak or inactive.

In rats the N-allyl isomers (Ia, $R = CH_2.CH:CH_2$) had no morphine-like properties but antagonized fentanyl-induced effects: the β -isomer (twice as active as nalorphine) was 4 times more effective than the α -form. In morphine-dependent monkeys the same derivatives precipitated a dose-related withdrawal syndrome with the β -isomer again the more effective agent.

Since certain N-allyl and N-cyclopropylmethyl 4-arylpiperidines (analogues of pethidine and its reversed ester and of ketobemidone) are analgesic "agonists" rather than antagonists (Casy, Simmonds & Staniforth, 1968; Oh-ishi & May, 1973), it is probable that, of the two classes, 3-arylpiperidines interact with opiate receptors in the more morphine-like manner. Flexible (boat) conformers of 3-arylpiperidines

			Molec.			Analysis	
1-substituent	Isomer	m.p. °	formula		С	H	N
(R in Ia)							
Me	α	248-2491	C14H22CINO	Found	65.78	8·77	5.49
				Required	65.74	8.66	5.47
Me	β	286-287 ²	C14H22CINO	Found	65.77	8.68	5.47
(CH ₂) ₂ Ph	à	215-217 ¹	$C_{21}H_{28}CINO$	Found	72.54	8.13	4.12
				Required	72.91	8·15	4.04
(CH ₂) ₂ Ph	β	224–225 ¹	$C_{21}H_{28}ClNO$	Found	71.19	8.23	3.90
	•		0.5 H ₂ O	Required	71.07	8.23	3.95
CH ₂ .CH:CH ₂	α	206-207 ²	C ₁₆ H ₂₄ ClNO	Found	68.42	8.62	5.00
			10 11	Required	68·18	8.58	4.96
CH ₂ .CH:CH ₂	β	233-235 ²	C ₁₆ H ₂₄ ClNO	Found	68·11	8.71	4.70

Table 1. 2,3-Dimethyl-3-m-hydroxyphenylpiperidine (Ia) hydrochlorides.

¹ From ethanol-ether.

² From ethanol.

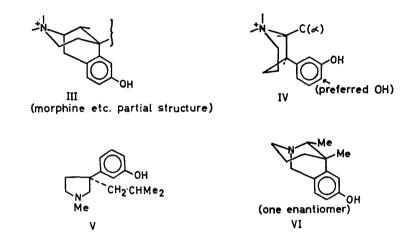
Table 2.	Nilsen and hot-plate	ED50 values in mice for	2,3-dimethyl-3-m-hydroxy-
	phenylpiperidines (Ia)	and related compounds.	

		ED50 mg kg ^{-1} , s.c.			
Compound ¹	Isomer	Nilsen method	Hot-plate method		
$Ia(R = CH_2CH_2Ph)$	α β	4·6 (2·5-8·5)) 11·3 (7·0-18·1)	7·0 (4·8–12·2) 10·6 (7·7–14·6)		
Ia(R = Me)	β	39.3 (22.7-68.2)	2		
VI		6.4 (3.1–13.1)	13.0 (8.8-19.1)		
Pethidine ³		3.5 (2.3-5.4)	4.7 (4.2-5.4)		

¹All hydrochlorides.

²Indeterminate.

^sPerrine & others (1972).



most clearly reveal the relation to the 3-dimensional skeleton of the morphine, morphinan, and benzomorphan classes of analgesic (III and IV). It is of interest that analgesically active derivatives of 3-arylpyrrolidines, e.g. V are converted to morphine antagonists when the ring nitrogen carries an allyl or cyclopropylmethyl substituent (Bowman, Collier & others, 1973a, b). These derivatives may also adopt conformations in which the basic and aromatic features are orientated similarly to those in flexible forms of 3-arylpiperidines. Such comparisons emphasize the necessity for a β -arylethyl moiety (present in all structural types discussed except 4-arylpiperidines) in analgesics capable of conversion to narcotic antagonists.

The 1-aza-des-*N*-6,7-benzomorphan VI, m.p. $210^{-}211^{\circ}$ from acetone–ether (Found: C, 76·46; H, 8·88; N, 6·54. C₁₄H₁₉NO requires: C, 77·38; H, 8·81; N, 6·45%), hydrochloride, m.p. $108^{-}109^{\circ}$ from ethanol–ether (Found: C, 61·81; H, 8·04; N, 5·17. C₁₄H₂₀ClNO·H₂O requires: C, 61·87; H, 8·15; N, 5·15%), derived from α -Ib (R = H) and formaldehyde-formic acid (Iorio & Casy, 1974) followed by *O*-demethylation, was about half and a third as active as pethidine in mice by Nilsen and hot-plate tests respectively (Table 2), and equi-active in rats by the tail withdrawal reflex.

We thank Dr. P. A. J. Janssen for testing the *N*-allyl isomers and VI in rats, Dr. E. L. May for accommodating one of us (M.A.I.) in his laboratory, and Drs L. Harris & M. Aceto (Virginia Medical College) for performing the antagonistic tests in monkeys.

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The isolation and enumeration of *Pseudomonas aeruginosa* from oily cream

The recovery of micro-organisms from topical non-sterile pharmaceuticals presents many problems for the analyst engaged in the microbiological quality control of such products. Variations of the membrane filtration technique developed by Sokolski & Chidester (1964) are widely used in the pharmaceutical and cosmetic industry to recover micro-organisms from topical preparations. Many solvents have been utilized to dissolve or disperse the ointment before membrane filtration. These include isopropyl myristate (IPM) (Sokolski & Chidester, 1964), n-hexane (White, Bowman & Kirshbaum, 1968), light liquid paraffin (British Pharmacopoeia, 1968) polyethylene glycol ether (Millipore Ltd., 1969), white spirit (Smith, personal communication) and carbon tetrachloride (Tall, personal communication). Recently Hambleton & Allwood (1972 & 1973) reported using a variety of solvents to recover *Bacillus megaterium* spores and *Escherichia coli* from white soft paraffin.

P. aeruginosa has been found as a common contaminant in topical preparations (Savin, 1967) and was isolated from oily cream by Simmons (1969). This report describes the recovery of *P. aeruginosa* from oily cream using a membrane filtration technique and a variety of solvent and dispersion systems.

P. aeruginosa NCTC 7244 was grown for 18h at 35° in nutrient broth (Oxoid Ltd., London) and found to contain approx. 5×10^8 cells ml⁻¹. The culture was diluted in sterile distilled water and 0·1 ml of a suitable dilution was incorporated into 2·0 g of oily cream by adequate mixing using a sterile spatula. The inoculated cream contained approx. 5×10^2 cells g⁻¹.

The following solvents were used in an attempt to recover *P. aeruginosa* cells from the cream; isopropyl myristate (Price's Chemicals Ltd., Bebington, Wirral, Cheshire),

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